

Effective from Session: 2020-21							
Course Code	BS441	Title of the Course	L	Т	Р	С	
Year	Ι	Semester	Ι	3	1	0	4
Pre-Requisite	UG in Biological Science	Co-requisite					
Course Objectives	various daily needs. The knowled	dge is used in health care	of general microbiology, contribution of microl e for prevention of diseases, diagnosis, steriliz od production, production of alcohol, in agricul	ation	method	s and d	lrug

Course	Course Outcomes						
CO1	Know about the history and development of Microbiology, taxonomy, genetic relationship						
CO2	Know about control and culturing of the microbes						
CO3	Know about the distinguished characteristics, morphology and importance of microbes						
CO4	Learn the preparation and use of culture media, Pure culture and cultural characteristics & preservation methods of microbes						
CO5	Know about the growth phases - kinetics, asynchronous, synchronous, batch and continuous culture						

Unit No.	Title of the Unit	Content of Unit	Contact Hrs.	Mapped CO			
1	History of Microbiology	8	CO-1				
2	Culture and control of microorganisms	Physical and chemical methods of control of microorganisms. Multidrug resistance in Microbes: importance & mechanisms. Microbial Culture techniques – principles and selective factors employed, enrichment systems –single cell isolation methods.	8	CO-2			
3	Diversity of microbes	Distinguished characteristics, general account on morphology, classification and economic importance of Algae, Protozoa and Fungi. Fungi as Plant Pathogens.	8	CO-3			
4	Bacterial Nutrition	Study of microbes - Preparation and use of culture media. Pure culture and cultural characteristics. Principles and methods of preservation of bacteria, yeasts and molds Bacterial Nutrition: Major nutritional types of bacteria, Microbial requirements of C, N, S, P, and microelements, growth factors, etc.	8	CO-4			
5	Growth Kinetics	Growth and control of microbes – Growth phases – kinetics, asynchronous, synchronous, batch and continuous culture. Factors affecting growth; Measurement of growth.	8	CO-5			
Refer	ence Books:						
1.	Gerherdt P, Murray	RG, Wood WH. Kreig, NR (1994) Methods for General and Molecular Bacteriology, ASM, Washington	n DC				
2.	Madigan MT, Mart	inko JM, Parker J. (1997) Biology of Microorganisms, Prentice Hall International Inc					
3.							
4.	4. Stanier RY, Ingraham JL, Wheelis ML, Painter PR (1992). General Microbiology, Mac Millan Education Ltd. London						
e-Le	earning Source:						

		Course Articulation Matrix: (Mapping of COs with POs and PSOs)										
PO- PSO CO	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PSO1	PSO2	PSO3	PSO4
CO1	3	1				2	2	1	3			
CO2	3	1					2	1	3			
CO3	3	1				2		1	3			
CO4	3	1				2		1	3		3	
CO5	3	1				2	2	1	3			

Name & Sign of Program Coordinator	Sign & Seal of HoD



Effective from Sess	sion: 2020-21						
Course Code	BS442	Title of the Course	le of the Course Biophysical methods L				
Year	Ι	Semester	Ι	3	1	0	4
Pre-Requisite	UG in Biological Science	Co-requisite					
Course Objectives	biotechnology-based researc	ch and industry. The co	e students with the understanding of various analyt urse will acquaint the students with the various instrun a generation and its analysis.				

		Course Outcomes
CO)1	To understand and learn various microscopy techniques used in biotechnology field.
CO	02	To understand and learn isolation of cellular fractions-separation, purification of proteins and amino acids, assay techniques for enzymes.
CO)3	Demonstrate principle and working of centrifugation and chromatography techniques.
CO	94	To learn the principles and applications of molecular techniques in microbiology.
CO	95	To learn principles and applications of various biophysical techniques used in the determination of biopolymer structures.

Unit No.	Title of the Unit	Content of Unit	Contact Hrs.	Mapped CO				
1	Містоѕсору	Principles and application of light phase contrast, fluorescence microscopy, scanning and transmission electron microscopy.	8	CO-1				
2	Isolation of cellular fractions	Separation, purification of proteins and amino acids, assay techniques for enzymes. Methods for lysis of plant, animal and microbial cell. Ultrafiltration, freeze drying and fractional precipitation. Use of detergents in isolation of membrane proteins.	8	CO-2				
3	Centrifugation	Ultracentrifugation - velocity and buoyant density determination. Density gradient centrifugation, molecular weight determination. Chromatography: Basic principles and applications of ion-exchange, types of ion- exchange resins, gel filtration, partition, affinity, HPLC and reverse phase chromatography, gas chromatography, TLC, Paper chromatography. Chromatofocussing.	8	CO-3				
4	Principles and applications of molecular techniques in microbiology	Electrophoresis: Agarose Gel electrophoresis, PAGE, Isoelectric focusing, capillary electrophoresis. Pulse field gel electrophoresis. RFLP, RAPD, ARDRA, RISA, Western, Northern and Southern blotting, FISH, Fluorescent activated cell sorting (FACS).	8	CO-4				
5	Determination of biopolymer structure	Principles and applications: X-ray diffraction, fluorescence, UV, visible, CD/ORD, ESR, NMR and Mass spectroscopy, Atomic Absorption Spectrophotometer, Plasma emission spectroscopy.	8	CO-5				
Referen	nce Books:							
1.	Protein Purification by	Robert Scopes, Springer Verlag Publication						
2.	1982 Tools in Biochem	istry David Cooper						
3.	Methods of Protein and Nucleic acid Research, Osterman Vol I – III							
4.	Centrifugation D. Rickwood							
5.	5. Practical Biochemistry, V th edition, Keth, Wilson and Walker							
e-Lea	rning Source:							

		Course Articulation Matrix: (Mapping of COs with POs and PSOs)										
PO- PSO	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PSO1	PSO2	PSO3	PSO4
СО	_			_		_		-			-	
CO1	3	1		3		3		3			3	
CO2	3	1		3		3		3			3	
CO3	3	1		3		3		3			3	
CO4	3	1		3		3		3			3	
CO5	3	1		3		3	1	3			3	

Name & Sign of Program Coordinator	Sign & Seal of HoD



Effective from Session: 2020-21							
Course Code	BS443	Title of the Course	Biomolecules	L	Т	Р	С
Year	Ι	Semester	Ι	3	1	0	4
Pre-Requisite	UG in Biological Science	Co-requisite					
	To understand the basics of	the understanding of bior	nolecules, the basic building blocks of living organ	isms, f	ocusing	on thei	r
Course Objectives	structural underpinnings, un	nique properties, biologica	l roles and functions and inter relations. Emphasis	will be	on the		
	association between structu	re and function of various	biomolecules at a chemical level with a biological	perspe	ctive.		

Course Outcomes						
CO1	The students will learn about carbohydrate types structure and functions.					
CO2	The students will learn about lipids: Definition and classification of lipids.					
CO3	The students will learn about proteins and amino acids structure, classification and functions.					
CO4	The students will learn about Nucleic acids (DNA and RNA), their composition, structure and functions					
CO5	The students will learn about water- and fat-soluble vitamins functions					

Unit No.	Title of the Unit	Content of Unit	Contact Hrs.	Mapp ed CO
1	Carbohydrates	Definition, classification, structure and functions of carbohydrates; Stereoisomerism, aldoses and ketoses; Important classes of monosaccharides, disaccharides, Structural and storage polysaccharides and mucopolysaccharides.	8	CO-1
2	Lipids	Lipids: Definition and classification of lipids. Nature of fatty acids. Role of triglycerides in energy storage and phospholipids in membrane formation, sterols, pigments.	8	CO-2
3	Proteins	Proteins: Nature of naturally occurring amino acids, Structure and functions of proteins (primary, secondary, tertiary and quaternary structure), Forces responsible for maintenance of protein structure	8	CO-3
4	Nucleic acids	. Nucleic acids: Composition of nucleic acids (ribo and deoxyribonucleic acids); Nucleosides, nucleotides and polynucleotides. Structure and function of DNA and RNA. Types of DNA: A, B and Z DNA, their structure and significance; Physical & biochemical properties of RNA: tRNA, rRNA, mRNA and hnRNA; Primary, secondary, and tertiary structures of RNA	8	CO-4
5	Vitamins:	Vitamins: Fat soluble and water soluble vitamins; elementary ideas about the physiological functions and deficiency diseases; Role of water soluble vitamins as co-enzyme precursor	8	CO-5
Refer	ence Books:			
1. Ec	kstein F, Lilley DM (1996). Catalytic RNA. Springer Verlag.		
2. Fre	eidberg EC, Walker C	C, Siede W. (1995). DNA Repair and Mutagenesis, ASM Press.		
3. Fre	eifelder D. (1991). M	olecular Biology. Narosa Publishing House		
4. Ga	rdener EJ, Simmons	MJ, Snustad DP. (1991). Principles of Genetics, John Wiley & Sons.		
e-Le	earning Source:			

		Course Articulation Matrix: (Mapping of COs with POs and PSOs)											
PO- PSO	PO1	DO1	DO2	PO4	PO5	PO6	PO7	PO8	PSO1	PSO2	PSO3	PSO4	
CO	POI	PO2	PO3	P04	POS	PU0	PO/	P08	P301	P302	P305	P304	
CO1	3	1						1		3			
CO2	3	1						1		3			
CO3	3	1						1		3			
CO4	3	1						1		3			
CO5	3	1						1		3			

Name & Sign of Program Coordinator	Sign & Seal of HoD



Effective from Session: 2	2020-21						
Course Code	BS444	Title of the Course	Microbial Cytology and Genetics	L	Т	Р	С
Year	Ι	Semester	Ι	3	1	0	4
Pre-Requisite	UG in Biological Science	Co-requisite					
Course Objectives		sm and regulation of euk	prokaryotic and eukaryotic cell organizatic aryotic cell cycle and signal transduction				

Course	Outcomes
CO1	The students will be able to explain prokaryotic cell organization, bacterial cell wall synthesis and details about antibiotics mechanism and
	development of antibiotic resistance.
CO2	The students will be able to describe eukaryotic cell organization, membrane function and transport, cytoskeletal elements and genetic
	organization.
CO3	The students will be able to discuss cell division in eukaryotes, cell cycle checkpoints and its regulation and various pathways of cell proliferation
	and apoptosis.
CO4	The students will be able to explain basics and mechanism of signal transduction, Quorum sensing and Biofilms.
CO5	The students will be able to explain methods of gene transfer in bacteria and different types of transposons present in prokaryotes.

Unit No.	Title of the Unit	Content of Unit	Contact Hrs.	Mapped CO	
1	Prokaryotic Cell Organization	Bacterial cell wall, Biosynthesis of peptidoglycan, basis of antibiotics, Mode of action of antibiotics, development of resistance, cytoplasmic membrane, ultrastructure of bacterial cell, Endospore, flagella, cell membrane, pili, capsule, prokaryotic genome.	8	CO-1	
2	Eukaryotic Cell Organization and protein targeting	Membrane biology: Structure, function, membrane protein transport in eukaryotes. Structure and functions of cell organelles, Cytoskeleton (structural proteins- microfilaments, actins, etc.), genetic organization (euchromatin, heterochromatin, Nucleosome model), concept of protein targeting.	8	CO-2	
3	Cell division and Eukaryotic Cell division cycle: Mitosis, Meiosis, Check points, role of cyclins and cyclin dependent kinases in its regulation. Cell proliferation and cell death, apoptosis.				
4	Cell Communication	8	CO-4		
5	Communication application. application. Gene transfer mechanisms in bacteria: Transduction: Generalized, restricted; Transformation Microbial Discovery, competence development, molecular mechanism of DNA uptake; Conjugation Genetics mechanism; mapping; Transposons in prokaryotes: Simple, composite, and complex transposon Mechanism of transposition; Retrotransposons. Simple, composite, and complex transposons		8	CO-5	
Refere	ence Books:				
1. A	lberts Bruce (1985) N	Iolecular Biology of Cell. Garland Pub			
2. C	onn Eric, Stumpf Pau	l K., Bruuening George, Doi Roy H., (1987) Outlines of Biochemistry Edition , John Wiley and Sons	, New Delhi	i.	
3. D	e Robertis E. D. P. an	d De Robertis E. M. F. (1987), Cellular and Molecular Biology Lea and Febiger, Philadelphia.			
4. S	chlegel Hans G. (199	5) General Microbiology, Edition 7, CUP, Cambridge.			
5. S	tanier R. Y., Adelberg	g E. A., Ingraham J. L., (1976) General Microbiology, 4th edition, Mac Millan Press, London.			
6. L	odish H, Berk A, Zipu	ursky SL et al. (2000) Molecular Cell Biology, 4th edn. New York: WH Freeman.			
7. Jo	oanne M. Willey, Lind	da M. Sherwood, Christopher J. Woolverton. Prescott's principles of microbiology, New York : McGra	aw-Hill, 201	12.	
e-Le	arning Source:				
t-Le	arming source.				

		Course Articulation Matrix: (Mapping of COs with POs and PSOs)											
PO- PSO	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PSO1	PSO2	PSO3	PSO4	
CO													
CO1	3	1				2	1	1		3			
CO2	3	1				2	1	1		3			
CO3	3	1				2		1		3			
CO4	3	1				2		1		3			
CO5	3	1				2		1		3			

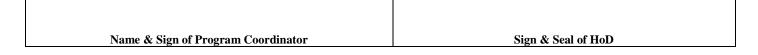
Name & Sign of Program Coordinator	Sign & Seal of HoD

Effective from Sessio	Effective from Session:											
Course Code	BS445	Title of the Course	Soil and Agricultural Microbiology	L	Т	Р	С					
Year	Ι	Semester	Ι	3	1	0	4					
Pre-Requisite	UG in Biological Science	Co-requisite										
Course Objectives	1 1 00	microbial diversity and the	lesigned with the objective to provide general intr e role of microorganisms in biogeochemical cycli									

	Course Outcomes								
CO1	Comprehend the physical, chemical and biological properties of soil and their importance.								
CO2	Have in depth knowledge of the role of microorganisms in plant growth particularly in rhizosphere and phyllosphere.								
CO3	Develop an understanding of the microbiology and physiology of C and N cycle specifically degradation of native and organic matter and								
	biological nitrogen fixation.								
CO4	Have knowledge of microbial transformation of elements as Phosphorus, Iron and Manganese								
CO5	Get insight of the types, production process, application methods and quality control of microbial biofertilizers.								

Unit No.	Title of the Unit	Content of Unit	Contact Hrs.	Mapped CO
1	Soil Microbiology	Structural and textural classes; Physico-chemical and biological properties of soil, soil enzymes, microorganisms and soil fertility. Methods used in soil chemistry and microbiological studies.	8	CO-1
2	Rhizosphere and Phyllosphere	Rhizosphere and Phyllosphere microorganisms, Rhizosphere effect, root exudates, influence of rhizosphere on crop productivity, plant growth promotingbacteria, biological control within microbial communities of rhizosphere, role of antibiotics and siderophore in biocontrol of plant pathogens, Induced resistance: Phytoalexins	8	CO-2
3	Biogeochemical cycles	8	CO-3	
4	Microbial transformation	Microbial transformation of Phosphorus, sulphur and micronutrients– Phosphorus cycle, mineralization of inorganic phosphates. Microbial transformation of Iron and Manganese.Microbial transformation of sulphur- Sulphur cycle, sulphur oxidizing and reducing microorganisms (Thiobacillus and Desulfovibrio).	8	CO-4
5	Biofertilizers	Definition and status of biofertilizer, types of biofertilizers. Nitrogenous and phosphatic biofertilizers - Rhizobium, Azotobacter, Azospirillum, Frankia, Vesicular Arbuscular Mycorrhiza and PSB/PSF Technologies for the production of biofertilizers. Methods of inoculation on seed and in soil. Quality control of biofertilizers.	8	CO-5
Referen	ce Books:			
1. Agri	icultural Microbiology -	- Rangaswami.		
2. Soil	Microbiology – Alxand	ler Martin.		
3. Soil	and soil microorganism	s – Subbarao		
e-Lear	rning Source:			
1. <u>h</u>	ttps://wachemo-elearnin	g.net/courses/agricultural-microbiology/#tab-course-section overview		

		Course Articulation Matrix: (Mapping of COs with POs and PSOs)											
PO- PSO CO	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PSO1	PSO2	PSO3	PSO4	
CO1	3	1					2	1	3		1		
CO2	3	1				2	2	1	3		1		
CO3	3	1					2	1	3	2			
CO4	3	1					2	1	3	2			
CO5	3	1			1	2	2	1	2		3		





Effective from Session: 2	2020-21						
Course Code	BS446	Title of the Course General Microbiology lab		L	Т	Р	С
Year	I	Semester	Ι	0	0	6	3
Pre-Requisite	UG in Biological Science	Co-requisite					
Course Objectives	regular basis in the biochemis	try labs i.e. tests for carbohyd form microbiology experimen	ctical hand on various biochemical assays rates, proteins, amino acids, cholesterol, its i.e. detection of gram positive and neg n in bacteria etc.	DNA a	and RN.	A. In	'n

Course	Course Outcomes								
CO1	Know the principles and instruments used in microbiology and various techniques.								
CO2	Students will learn pure culture techniques and know how to enumerate microbes from soil samples.								
CO3	Know how to perform Gram staining, spore staining for bacteria, and fungal staining followed by microscopic examination and biochemical								
	identification of bacteria.								
CO4	Students will know how to determine bacterial motility and isolate Rhizobium from nodules.								
CO5	Students will also be able to perform biochemical estimations of macromolecules in a given sample.								

Exp. No.	Title of Experiment	Contact Hrs.	Mapped CO
Exp-01	General instructions, Microbiology laboratory and its discipline.	6	CO-1
Exp-02	Handling of microscopes, Calibration and measurement of microscopic objects.	6	CO-1
Exp-03	Cleaning of glassware and sterilization. Preparation and use of glassware cleaning solutions, sterilization.	6	CO-1
Exp-04	Pure culture techniques: serial dilution, pour plate, spread plate, streak plate methods	6	CO-2
Exp-05	Enumeration of bacteria, fungi and actinomycetes from soil samples.	6	CO-2
Exp-06	Culture and microscopic examination of bacteria by staining methods - Gram's, capsule and spore staining.	6	CO-3
Exp-07	Culture and microscopic examination of fungi by Lacto-phenol cotton blue staining.	6	CO-3
Exp-08	Identification techniques: morphological and biochemical identification of bacteria using Bergey's Manual of Determinative Biology	6	CO-3
Exp-09	Motility of bacteria.	6	CO-4
Exp-10	Isolation of Rhizobium from nodules	6	CO-4
Exp-11	Estimation of carbohydrates, protein, DNA, RNA, and chlorophyll	6	CO-5
Referenc	e Books:		L
1. Ca	ppuccino, J. C. and Sherman, N. (1992). Microbiology: A laboratory manual, Addison Wesley Pub. Co		
2. Be	nson HJ (1994). Microbiological Applications, WmC Brown Publishers, Oxford.		
3. Co	llins C.H, Lyne P.M, (1985). Microbiological methods. Butterworths, London.		
4. Rh	odes P.M, Stanbury P.F. Applied Microbial Physiology - A practical approach. IRL Press, Oxford University Press	, Oxford.	
5. Wi	lson K, Walker J. (1995) Practical Biochemistry Principles and Techniques, Cambridge University Press		
6. K.I	R. Aneja Bergey's Manual of Determinative Bacteriology		
e-Learı	ing Source:		
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	Course Articulation Matrix: (Mapping of COs with POs and PSOs)											
PO- PSO	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PSO1	PSO2	PSO3	PSO4
СО												
CO1	3	3	1			3		3	1		3	2
CO2	3	3	1			3	1	3	3		3	2
CO3	3	3	1			3		3	3		3	2
CO4	3	3	1			3		3	3		3	2
CO5	3	3	1			3		3		3	3	2

Name & Sign of Program Coordinator	Sign & Seal of HoD



Effective from Sessio	m: 2020-21						
Course Code	BS451	Title of the Course Microbial Metabolism		L	Т	Р	С
Year	Ι	Semester	II	3	1	0	4
Pre-Requisite	UG with Biological Science	Co-requisite					
Course Objectives	metabolism and pathway analys	is. It also gives understant biomolecules. The co	evelop an understanding of catabolism, anabolisn anding of how enzymes and metabolites in living urse also imparts comprehensive knowledge abou protein, lipid and nucleic acid.	systen	n work 🕯	to produ	

Course	Outcomes
CO1	Understand the concept of enzymes and enzyme kinetics
CO2	Comprehend the carbohydrate metabolism, significance of glycolysis and ETC
CO3	Acquire knowledge about the metabolism of lipids, amino acids and nucleic acids.
CO4	Understand the basics of microbial degradation of Xenobiotics and Fermentation: Special pathways for primary attack on organic compounds
	by microorganisms
CO5	Have knowledge about the Nitrogen metabolism and Biological nitrogen fixation

Unit No.	Title of the Unit	Content of Unit	Contact Hrs.	Mapped CO	
1	Enzymes	Classification, properties and factors influencing enzyme activity, co-enzymes, prostheticmesgroup and co-factors, Lock & key hypothesis, induced fit hypothesis, Enzyme kinetics:Michelis Menten equation, Lineweaver-Burk plot, Enzyme inhibition, Allosteric enzymes			
2	Aerobic and anaerobic metabolism in bacteriaAerobic and anaerobic metabolism in bacteria - role of ATP, reducing powers and Biochemistry of catabolic reactions in aerobic heterotrophs: Glycolysis, hexose monophosphate shunt and Entner doudoroff pathways, TCA cycle, Role of glyoxylate cycle in acetic acid oxidation. Electron transport chain and oxidative phosphorylation, Gluconeogenesis8		8	CO-2	
3	Metabolism of lipids, amino acids and Nucleic acids Oxidation of fatty acid (beta-oxidation) and its biosynthesis. Metabolism of amino acids. Biosynthesis and degradation of nucleotides		8	CO-3	
4	Microbial degradation of Xenobiotics and Fermentation Special pathways for primary attack on organic compounds by microorganisms, Catabolic reactions of anaerobic chemohetrotrophs, Anaerobic respiration and fermentation. Autotrophic nutrition of microorganisms. Bacterial photosynthesis		8	CO-4	
5	Nitrogen metabolism	Biological nitrogen fixation: nitrogenase enzymes, structure and properties, nif' gene: regulation and functions. Physiology and biochemistry of nitrogen fixation, denitrification, nitrate and nitrite reduction, sulphate and sulphur reduction, H2S formation, deamination and transamination. Utilization of various nitrogen sources (ammonia, urea, nitrate, amino acids) by bacteria.	8	CO-5	
Referen	nce Books:				
1. Bro	ck —Biology of Microc	organisms			
2. Bro	wn, T.A. —Gene cloning	g: An introduction			
3. Frei	felder, DM —Molecula	r Biology			
4. Leh	ninger —Biochemistry				
5. Levi	ine- Genes				
e-Lea	rning Source:				

		Course Articulation Matrix: (Mapping of COs with POs and PSOs)										
PO- PSO	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PSO1	PSO2	PSO3	PSO4
CO	POI	PO2	POS	P04	POS	POo	PO/	P08	P301	P302	PS05	P304
CO1	3	1						1		3		
CO2	3	1						1	3	3		
CO3	3	1						1		3		
CO4	3	1					1	1	3	3		
CO5	3	1					2	1	3	3		
	•	•	1- Low (Correlation:	2- Moderat	e Correlatio	n: 3- Subst	antial Corr	elation	•	•	



Effective from Sessio	Effective from Session: 2020-21							
Course Code	MT412	Title of the Course	Bioinformatics and Biostatistics	L	Т	Р	С	
Year	Ι	Semester	П	3	1	0	4	
Pre-Requisite	UG in Biological Science	Co-requisite						
Course Objectives	To understand the basics of	the computer bioinfor	matics and statistical analysis					

Course	Course Outcomes					
CO1	To understand the Basics of computers.					
CO2	To understand the biological data formats					
CO3	To understand the mechanisms of sequence analysis.					
CO4	To understand the biostatistics					
CO5	To understand the correlation analysis					

Unit No.	Title of the Unit	Content of Unit	Contact Hrs.	Mapped CO						
1	Basics of computers	Block diagram of computer, input and output devices, storage devices, operating systems – DOS, Windows, Linux. Basics of networking and their types, topologies, INTERNET: TCP/IP, World Wide Web, e-mail etc.	8	CO-1						
2	Biological data file formats	*.FASTA, *.PIR, *.GDE, *.PDB, Alignment files (*.ALN) etc. Search engines: ENTREZ, DBGET, SRS etc. Primary nucleotide sequence atabases: Genbank, EMBL, DDBJ; Primary Protein sequence databases: SwissProt, Protein information resources, TREMBL. Etc. Secondary databases: PROSITE, PRINTS, BLOCKS, PFAM.; Microbiology DATABASES: ICTV, Animal Virus Information System (AVIS).	8	CO-2						
3	Sequence analysis Needleman wunsch, Smith Watermann algorithms, Sequence similarity search programs – BLAST and FASTA. Substitution matrices: PAM, BLOSSUM. Multiple sequence alignments: Center Star		8	CO-3						
4	Biostatistics	Measures of central tendency – mean (arithmetic, harmonic & geometric) median and mode: Measures		CO-4						
5	Correlation analysis	Positive and negative correlation, Karl Pearson's coefficient of correlation, Spearman's rank correlation. Regression analysis: regression line Y on X and X on Y, angle between two regression lines. Test of significance: null and alternative hypothesis, level of significance, Z-test, Student_t'-test, Chi-square test for goodness of fit and independence of attributes.	8	CO-5						
Refer	ence Books:									
1. D	eveloping Bioinforr	natics Computer Skills: Cynthia Gibas & Per Jambeck – 2001								
2. SI	hroff Bioinformatics	s Basics: Applications in Biological Science and Medicine – 2002								
3. H	H Rashidi & LK Bu	hehler, CRC Press, London Bioinformatics: Sequence, structure and databanks – 2000								
4. E	Des Higgins & Willi	e Taylor – Bioinformatics: A practical guide to the analysis of genes and proteins – 2001								
5. A	D Baxevanis & BF	F Ouellette – Wiley Interscience – New York								
e-Le	earning Source:									

		Course Articulation Matrix: (Mapping of COs with POs and PSOs)										
PO- PSO	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PSO1	PSO2	PSO3	PSO4
CO												
CO1	3	1				2		1		3	2	
CO2	3	1				2		1		3	2	
CO3	3	1				2		1		3	2	
CO4	3	1				2		1				3
CO5	3	1				3		1				3

Name & Sign of Program Coordinator	Sign & Seal of HoD



Effective from Session: 2020	0-21						
Course Code	BS452	Title of the Course	Molecular Biology	L	Т	Р	С
Year	Ι	Semester	II	3	1	0	4
Pre-Requisite	UG in Biological Science	Co-requisite					
To develop in students the understanding about the molecular biology of the microbes and detailed knowledge of mo							
Course Objectives	mechanism of gene expression and	its regulation.					

	Course Outcomes							
CO1	The students will be able to explain the detailed mechanism of DNA replication and regulation in prokaryotes and eukaryotes.							
CO2	The students will be able to discuss the characteristics of promoter and mechanism of transcription in prokaryotes and eukaryotes.							
CO3	The students will be able to explain the detailed mechanism of translation and its regulation in prokaryotes and eukaryotes.							
CO4	The students will be able to describe in detail the types of post-transcriptional and post translation modifications in eukaryotes.							
CO5	The students will be able to explain the regulation of gene expression in different organisms and methods of DNA repair.							

2 Tr	ONA replication	DNA replication: Origin of replication; Mechanism of DNA replication (initiation, elongation and termination); Roles of DNA polymerases and other proteins involved in replication; Replication in eukaryotes. Fidelity and regulation of replication. σ or Rolling circle replication in ϕ X174. Transcription: Mechanism of transcription in prokaryotes and eukaryotes (initiation, elongation and termination); RNA polymerases: structure, subunits and function. Promoter; Transcription factors; Enhancer and other regulatory elements of eukaryotes. Reverse transcription. Translation in prokaryotes and eukaryotes: Adapter role of tRNA, Evidence for a triplet code; Properties of Genetic code; Wobble hypothesis; A, P and E sites of ribosome; Ribosome binding site; Formation of initiation complex; Ribosome cycle; Initiation, elongation and termination of translation in prokaryotes and eukaryotes. Roles of Initiation factors, Elongation factors, Release	8	CO-1 CO-2
		and termination); RNA polymerases: structure, subunits and function. Promoter; Transcription factors; Enhancer and other regulatory elements of eukaryotes. Reverse transcription. Translation in prokaryotes and eukaryotes: Adapter role of tRNA, Evidence for a triplet code; Properties of Genetic code; Wobble hypothesis; A, P and E sites of ribosome; Ribosome binding site; Formation of initiation complex; Ribosome cycle; Initiation, elongation and termination of translation in prokaryotes and eukaryotes. Roles of Initiation factors, Elongation factors, Release		CO-2
3 Tr	ranslation	Properties of Genetic code; Wobble hypothesis; A, P and E sites of ribosome; Ribosome binding site; Formation of initiation complex; Ribosome cycle; Initiation, elongation and termination of translation in prokaryotes and eukaryotes. Roles of Initiation factors, Elongation factors, Release	8	
		factors, Aminoacyl tRNA synthetase	*	CO-3
tra 4 an tra	ost – ranscriptional nd post- ranslational nodifications	Post - transcriptional / Co-transcriptional processing of rRNA, mRNA, tRNA: Addition of 5' cap and 3' Poly A tail in mRNA, RNA splicing - Self splicing and Spliceosome mediated splicing, Alternative splicing; Cutting events or action of ribonucleases, Covalent modifications, RNA editing. Post-translational processing: Intein splicing, Chemical modification, Proteolytic cleavage, Zymogen activation; Protein degradation Ubiquitin-Proteosome Pathway; Polycistronic and monocistronic. Inhibitors of transcription and translation.	8	CO-4
	egulation of ene expression	Regulation of gene expression: Concept of operon: Lac and Trp operons, Eukaryotic gene expression, Significance of repressor, Attenuation; histone modifications, Mutation: Spontaneous, induced; Chemical and physical mutagens; Nonsense mutation; Missense mutation; Frame shift mutation; Suppressor mutation, DNA repair mechanisms: Photoreactivation, Base excision repair, Nucleotide excision repair, Mismatch repair, Recombination repair, Translesion DNA synthesis.	8	CO-5
Reference	Books:			
		s VII. Oxford University press.		
		Berk A, Zipursky SL, Darnell J. (1995). Molecular cell biology.		
		H, Roberts JW, Steitz JA, Weiner AM. (1987). Molecular biology of the gene.		
	e :	f Biochemistry (2017) by Nelson and Cox Seventh edition, WH Freman and Co.		
5. Voet	t, Donald, and Judit	th G. Voet. Biochemistry. New York: J. Wiley & Sons, 1995. Print		
e-Learnin	ng Source:			

https://www.youtube.com/watch?v=TNKWgcFPHqw

			Course Ar	ticulation N	latrix: (Maj	oping of CO	s with POs	and PSOs)			
PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PSO1	PSO2	PSO3	PSO4
3	1				1		1		3		
3	1				1		1		3		
3	1				1		1		3		
3	1				1		1		3		
3	1				1		1		3		
	PO1 3 3 3 3 3 3 3 3	PO1 PO2 3 1 3 1 3 1 3 1 3 1 3 1 3 1 3 1 3 1	PO1 PO2 PO3 3 1						PO1 PO2 PO3 PO4 PO5 PO6 PO7 PO8 PS01 3 1 1		

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Effective from	Session:						
Course Code	BS453	Title of the Course	Industrial Microbiology & Fermentation Technology	L	Т	Р	С
Year	Ι	Semester	II	3	1	0	4
Pre-Requisite	UG in Biological Science	Co-requisite					
Course Objectives	improved biochemical or ph	ysiological fermentatio	e to develop an understanding of Industrial microbiology n are mainly carried out by fungi and bacteria on large scal tion is to produce highest quality and quantity of particles p	e to pi	oduce c	commer	cial

	Course Outcomes						
CO1	Know the basics of fermentation technology.						
CO2	Have insight to the general design of fermenter, media and the process of fermentation						
CO3	Understand the relation between growth and product formation, optimization of fermentation process and DSP.						
CO4	Have knowledge of how microbes are used for production of important industrial products.						
CO5	Have basic knowledge of intellectual property rights specially patents.						

Unit No.	Title of the Unit	Content of Unit	Contact Hrs.	Mapped CO			
1	Introduction to Industrial Microbiology	fermentation (Batch and continuous). Fermentation media-Types of fermentation media, sources of carbon, nitrogen, trace elements, growth factors, precursors, buffers, antifoam agents, sterilization of media					
2	General design of fermenter, concept and importance of gas exchange and mass transfer and scale-up in microbial fermentation. Processes of fermentation. Basic concept of cell and enzyme immobilization and reactors used for immobilized enzymes		8	CO-2			
3	3 Growth and product formation Growth and product formation: Definition of primary and secondary metabolites, and their products. Overproduction of industrially important metabolites by strain improvement; Product recovery and techniques involved in downstream processing		8	CO-3			
4	4Microbial production of industrially important productsA brief idea about the products obtained from microbes, commercial production of citric acid and glutamic acid, antibiotics (as penicillin), solvents (ethanol), vitamins (B12), enzymes (Protease). Production of single cell protein- Microorganisms and substrates used, techniques of production, merits and demerits of single cell protein.						
5	Intellectual property rights	Introduction to intellectual property rights; Intellectual property laws; significance of IPR. Forms of IPR like patent, design copyright and trademark. Requirement of a patentable novelty; Issues related to IPR protection of software and database; IPR protection of life forms. Obtaining patent; Invention step and prior art and state of art procedure; Detailed information on patenting biological products and biodiversity. Trade related aspects of Intellectual Property Rights and Budapest treaty.	8	CO-5			
Referen	ce Books:						
1. Prir	nciples of fermentation	technology by P. Stanbury & Allan Whitekar, Pergamon					
2. Pre	ss Industrial microbiolo	gy by Cruger and Cruger W. Sinauer Associates; Madison,					
3. Ind	ustrial Microbiology by	L.E Casida , John Wiley and sons INC.					
4. Pre	scott and Dunn,s Indust	ral microbiology, 4th edition (1982) by Gerald Reed.					
e-Learn	ing Source:						
1. http	os://onlinecourses.nptel.	ac.in/noc19_bt20/preview					
2. http	os://onlinecourses.swaya	m2.ac.in/cec22_bt18/preview					

				Course Ar	ticulation N	latrix: (Maj	oping of CO	s with POs	and PSOs)			
PO- PSO	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PSO1	PSO2	PSO3	PSO4
CO CO1	3	1				1	1	1	3			
CO2	3	1				1		1			3	
CO3	3	1				1		1	1		3	
CO4	3	1				2		1	1		3	
CO5	3	1		3	3	2	1	1				3

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Effective from Session	Effective from Session:								
Course Code	BS454	Title of the Course	Microbial Diversity	L	Т	Р	С		
Year	Ι	Semester	II	3	1	0	4		
Pre-Requisite	UG in Biological Science	Co-requisite							
Course Objectives	Course Objectives On completion of this course, students will be able to develop an understanding of microbial diversity								

Course	Course Outcomes						
CO1	Microbial ecology – concepts of Niche, habitat, ecosystem etc.						
CO2	Microbial interactions: symbiosis, synergism, fungal and algal association with plants.						
CO3	General characteristic of purple and green sulphur bacteria, Cyanobacteria and Prochlorales, BGA in agriculture.						
CO4	Methanogenic Archeobacteria—General characteristics. Bioluminescent and nitrogenfixing bacteria. Magnetotactic bacteria						
CO5	Microorganisms in prospecting of oils Extremophiles- Acidophilic, alkalophilic, psychrophilic, thermophilic and halophilic microorganisms						

Unit No.	Title of the Unit	Content of Unit	Contact Hrs.	Mapped CO
1	Microbial ecology	Concept of habitat and ecological niches, Ecosystem, Energy flow, food chain, food web, biotic community concept, Microbial succession, adaptation and natural selection of microbial population.	8	CO-1
2	Microbial interactions	Symbiosis, Synergism, Commensalism, Ammensalism, Predation and Parasitism, Mycorrhizal associations-structure, characteristics and their role in Agriculture and Forestry, Algal association with other microorganisms and plants	8	CO-2
3	Photosynthetic microbes	Anoxygenic photosynthetic microbes General characteristic of purple and green sulphur bacteria. Oxygenic photosynthetic microbes- General characteristics of Cyanobacteria and Prochlorales; Role of blue green algae (BGA) in agriculture	8	CO-3
4	Archeobacteria	Methanogenic Archeobacteria—General characteristics. Bioluminescent and nitrogenfixing bacteria- A high energy spending bacteria. Magnetotactic bacteria Microorganisms in prospecting of oils Extremophiles- Acidophilic, alkalophilic, psychrophilic, thermophilic and halophilic microorganisms.	8	CO-4
5	Microbes of toxic environments and Biodeterioration	Acid mine drainage, coal desulphurisation, waste containing cyanides, xenobiotics, pesticides and chemicals, heavy metals, hydrocarbons & radio isotopic materials Concept of autotrophy – an example of extreme synthesis Biodeterioration-concept, biodeterioration of wood, stonework, pharmaceutical products, rubber, plastic,	8	CO-5
Referen	ce Books:			
1. Ext	remophiles-(2000) By E	3.N.Johari Springer Verlag,New York.		
2. Mic	crobial diversity (1999)	by D.Colwd Academic press. and Wilkins, Baltimore Academic press		
3. Ber	gy's Manual of Systema	atic Bacteriology (1984). Vols. I and III . Williams and Wilkins, Baltimore Academic press		
e-Lear	ning Source:			
	0			

		Course Articulation Matrix: (Mapping of COs with POs and PSOs)										
PO- PSO	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PSO1	PSO2	PSO3	PSO4
CO												
CO1	3	1					3	2	3			
CO2	3	1					3	2	3			
CO3	3	1					3	2	3			
CO4	3	1					3	2	3			
CO5	3	1					3	2	3			
		•	1 Low (appolation	2 Moderat	o Correlatio	n. 3. Subst	ontial Corr	lation	•	•	

1- Low Correlation; 2- Moderate Correlation; 3- Substantial Correlation

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Effective from Session: 2020-2021										
Course Code	BS455	Title of the Course	Mycology and Plant Microbe Interactions	L	Т	Р	С			
Year	Ι	Semester	II	3	1	0	4			
Pre-Requisite	UG in Biological Science	Co-requisite								
The objective of this course is to develop an understanding of the fungi, lichen and interaction of microbes to pla										
Course Objectives	understand different plant di	understand different plant diseases caused by fungi								

	Course Outcomes
CO1	Students will gain insight into the general characters of fungi, their nutritional types and genetic variation.
CO2	Students will acquire knowledge of the general classification and main groups of fungi.
CO3	Comprehend the economic importance of fungi, the biology and importance of lichens and the role of saprotrophs in ecosystems.
CO4	Students will develop basic understanding of the complex plant microbe interaction in Rhizosphere and phyllosphere and know the
	microorganisms acting as biofertilizers and biopesticides or causing diseases and understand the factors influencing plant diseases.
CO5	Students will have knowledge of some common Plant Diseases: including their epidemiology and symptoms.

Unit No.	Title of the Unit	Content of Unit	Contact Hrs.	Mapped CO				
1	Fungi	Historical account; General characters of fungi with special reference to thallus organization and reproduction in fungi. Nutritional types of fungi: biotrophs, hemibiotrophs, symbionts and necrotrophs and life cycle in fungi. Genetic variation in fungi- heterocaryosis and parasexual cycle and their significance. Sex hormones in fungi.	8	CO-1				
2	General classification of fungi	Study of the following main groups of fungi: Myxomycota with special reference to Stemonitis; Plasmodiophormycetes with special reference to Plasmodiophora; Oomycetes with special reference to Pythium.; Zygomycotina with special reference to Zygorhynchus; Ascomycotina with special reference to Yeasts, Protomyces, Aspergillus, Taphrina; Basidiomycotina with special reference to Puccinia, Agaricus; Deuteromycotina with special reference to Alternaria.	8	CO-2				
3	Economic importance of fungi	Lichens: types, biology and physiology of lichen thallus, economic importance of lichens; Mycorhiza. Beneficial uses of fungi, industrial production of enzymes and penicillin. Edible Mushrooms. Fungi as animal parasites, mycoses of vertebrates types and symptoms. Insect fungus association. Role of saprotrophs in ecosystems.	8	CO-3				
4	Plant Microbe interaction	Interaction of microbes in Rhizosphere and phyllosphere. Plant growth promotion and its mechanisms, Biofertilizers and biopesticides. Plant pathogens: Koch's postulates. Classification of plant diseases. Dissemination of phytopathogens. Causal agents of plant diseases. General symptoms of plant diseases. Factors influencing infection, colonization and development of symptoms. Specialization of parasitism, pathogenesis: role of enzymes and toxins in pathogenesis. Genetics of host- pathogen interaction. Defense mechanism in host: effect of infection on host physiology. Control of plant pathogens (plant quarantine; Cultural, Physical, chemical & biological methods of control).	8	CO-4				
5	Plant Diseases	Epidemiology, symptoms, etiology, perennation and control of following diseases: Damping off of seedling and fruit rot- Pythium; Stem gall of coriander-Protomyces macrospores; Peach leaf curl-Taphrina deformans; Rust of wheat- Puccinia recondite; Covered smut of barley-Ustilago hordei; Leaf spot and shot holes- Alternaria spp. Citrus canker; Tobacco mosaic disease; Root knot of vegetables-Meloidogyne; Abiotic/Non pathogenic diseases – Black tip of mango; Mycotoxins and storage diseases.	8	CO-5				
Referen	ce Books:							
1- Aneja	, K.R. & Mehrotr	a, R.S. (2011). Fungal Diversity & Biotechnology. New Age International Publishers, New Delhi.						
2- Alexo	poulos, C. J., Mir	ns, C.W. and Blackwell, M. (1996). Introductory Mycology. 4th edition John Wiley & Sons, USA.						
3- Mehro	otra, R.S. and And	eja, K.R. (2010). Introduction to Mycology. Wiley Eastern Ltd. New Delhi.						
4- Moor	4- Moore –Landcker, E. (1996). Fundamentals of the Fungi. Prentice Hall.							
5- Agrio	5- Agriose, G.N. (2005). Plant Pathology, 5th edition Academic Press, Inc., Ainsworth, G.C. and Sussman, A.A. (Eds).							
6- Deaco	on J.W. (1997). M	lodern Mycology (Basic Microbiology) 3 rd Ed. Wiley Blackwell.						
e-Lear	ning Source:							

	Course Articulation Matrix: (Mapping of COs with POs and PSOs)											
PO- PSO	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PSO1	PSO2	PSO3	PSO4
CO												
CO1	3	1					3	1	3			
CO2	3	1					3	1	3			
CO3	3	1				2	3	1	3			
CO4	3	1			1	1	3	1	3			
CO5	3	1			1	1	3	1	3			
			1- Low	[·] Correlation	n; 2- Modera	te Correlati	on: 3- Sub	stantial Co	orrelation			

1º Low Correlation, 2º Woder at Correlation, 3º Substantial Correlation

Name & Sign of Program Coordinator	Sign & Seal of HoD



Effective from Session	Effective from Session: 2020-21							
Course Code	BS412	Title of the Course	Enzymology & Enzyme Kinetics	L	Т	Р	С	
Year	Ι	Semester	Ι	3	1	0	4	
Pre-Requisite	UG in Biological Science	Co-requisite						
Course Objectives	This course has been designed to teach the student majoring in science all the major aspects of the study of enzymes. The course focuses on the theories of enzyme kinetics, the mechanisms of enzyme catalysis, and immobilization of enzyme.							

	Course Outcomes
CO1	The students will understand the general properties of enzymes and their classification & nomenclature.
CO2	The students will understand the theories of enzyme kinetics.
CO3	The students will understand the mechanisms of enzyme catalysis and enzyme inhibition & activation.
CO4	The students will understand the Multisubstrate enzyme kinetics.
CO5	The students will understand the enzyme Immobilization and its clinical & industrial use.

Unit No.	Title of the Unit	Content of Unit	Contact Hrs.	Mapped CO
1	Classification and nomenclature of enzymes	General properties of enzymes. Mechanism of enzyme action: Chymotrypsin, ribonuclease, activation of transition metal cation, activation by alkaline earth metal cation, nicotinamide nucleotide, flavin nucleotide and adenosine phosphate.	8	CO-1
2	Enzyme kinetics	Michaelis-Menten initial rate equation based on equilibrium assumption, Briggs Haldane steady state approach, integrated form of the Michaelis equation, methods for the determination of Km and Vmax normalized initial rate equation and normalized curves, Haldane relationship.	8	CO-2
3	Effect of factors and inhibitors on enzyme kinetics	Effect of enzymes concentration, pH and temperature on kinetics of enzyme reactions. Enzyme inhibition and activation: Types of reversible inhibitors, qualitative analysis of data, derivation of equations for different types of inhibitions, determination of inhibitor constant, determination of activator constant.	8	CO-3
4	Multisubstrate enzyme kinetics	Multisubstrate enzyme kinetics: random bi-bi, and ping pong reactions. Intracellular localization of enzymes, purification of enzymes and tests for homogeneity.		CO-4
5	Applied Enzymology	Immobilization; kinetics of immobilized systems. Isozymes. Allosteric enzymes. Industrial and clinical scope of enzymes.	8	CO-5
Referen	ce Books:			
1. Enzy	ymes Biochemistry, H	Biotechnology, Clinical Chemistry Authors: T Palmer, P L Bonner; Woodhead Publishing		
2. Bioc	chemistry – Lubert St	ryer Freeman International Edition.		
3. Lehr	ninger: Principles of l	Biochemistry (2017) by Nelson and Cox Seventh edition, WH Freman and Co.		
4. Enzy	yme Structure and M	echanism; Publisher W H Freeman & Co, New York; Alan Fersht		
5. Enzy	ymes: Authors: Malco	olm Dixon, Edwin C. Webb; Academic Press		
e-Lear	ning Source:			

				Course A	Articulation N	Matrix: (Ma	pping of C	Os with P	Os and PSOs)		
PO- PSO	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PSO1	PSO2	PSO3	PSO4
СО												
CO1	3	1				2		1		3		
CO2	3	1				2		1		3		
CO3	3	1				2		1		3		
CO4	3	1				2		1		3		
CO5	3	1				2		1		3	3	

1- Low Correlation; 2- Moderate Correlation; 3- Substantial Correlation

Name & Sign of Program Coordinator

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Effective from Sess	Effective from Session: 2020-21									
Course Code	BS456	Title of the Course	Applied Microbiology and Bioinformatics Lab	L	Т	Р	С			
Year	Ι	Semester	Semester II		0	12	6			
Pre-Requisite	UG in Biological Science									
Course Objectives	phyllosphere/rhizosphere micro testing using bacterial system. I	bial flora, detection of e Basics of computers – basics and along with the unders	factors involved in bacterial growth along with Enu xtracellular microbial enzymes and antibiotic sensit asic commands – file creation, copying, moving & c standing of various biological databases. The provid lysis and gene prediction	ivity a leletin	nd/or to g in DO	DS &				

	Course Outcomes
CO1	Measurement of bacterial growth/growth curve, Effect of physical and chemical factors on the growth of bacteria: temperature, pH, and
	salts and Enumeration of phyllosphere/rhizosphere microbial flora and Enumeration/Isolation of PSB/PSF
CO2	Detection of extracellular microbial enzyme: Beta lactamases, Testing for antibiotic sensitivity and/or toxicity using bacterial system,
	Determination of MIC values (tube dilution and spot plate method), Screening for antibiotic producing microbes and Microbiological
	examination of milk and milk products
CO3	Microbiological quality testing of milk (MBRT test) and Microbial examination of industrial waste water/sewage.
CO4	Understanding basics of computers - basic commands - file creation, copying, moving & deleting in DOS & Windows. Internet - Using
	browsers - search engines and understanding use of various biological databases-GENBANK, EMBL, Swissprot - Protein Data Bank
CO5	Performing different types of sequence analysis queries in BLAST and FASTA. (Homology search), Multiple sequence alignments
	(Clustal) and Phylogenetic Analysis. (Phylip or Clustal) and Gene Prediction

Exp. No.	Title of Experiment	Contact Hrs.	Mapped CO
Exp-01	-01 Measurement of bacterial growth/growth curve, Effect of physical and chemical factors on the growth of bacteria: temperature, pH, and salts		CO-1
Exp-02	p-02 Enumeration of phyllosphere/rhizosphere microbial flora. Enumeration/Isolation of PSB/PSF		CO-1
Exp-03	p-03 Detection of extracellular microbial enzyme: Beta lactamases, Testing for antibiotic sensitivity and/or toxicity using bacterial system, Determination of MIC values (tube dilution and spot plate method)		CO-1
Exp-04	Screening for antibiotic producing microbes	6	CO-2
Exp-05	p-05 Microbiological examination of milk and milk products, Microbiological quality testing of milk (MBRT test).		CO-3
Exp-06	Microbial examination of industrial waste water/sewage.	6	CO-4
Exp-07	Basics of computers – basic commands – file creation, copying, moving & deleting in DOS & Windows. Internet - Using browsers – search engines	6	CO-3
Exp-08	Using biological databases – GENBANK, EMBL, Swissprot – Protein Data Bank.	6	CO-4
Exp-09	Different types of sequence analysis queries in BLAST and FASTA. (Homology search).	6	CO-4
Exp-10	Multiple sequence alignments (Clustal) and Phylogenetic Analysis. (Phylip or Clustal)	6	CO-5
Exp-11	Gene Prediction.	6	CO-5
Reference	ee Books:		
Mi	rhardt P. Murray RG, Wood WA, and Kreig NR (ed.) (1994) Methods for General and Molecular Bacteriology crobiology, Washington D.C.	-	
	rick R. Murray. (editor chief) (1999) Manual of clinical microbiology, 7 Th edition, ASM Press, Washington 198) Pathological techniques - Anmol Publications Pvt. Ltd. N.D.	D.C. • Prakash M., A	Arora, C.K.
3. Sai	nbrook J, Fritsch EF, Maniatis T. (1989). Molecular coloning. Cold Spring Harbor Laboratory Press.		

e-Learning Source:

		Course Articulation Matrix: (Mapping of COs with POs and PSOs)										
PO- PSO	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PSO1	PSO2	PSO3	PSO4
CO												
CO1	3	3	1			3	3	3	3		3	2
CO2	3	3	1			3	3	3	3		3	2
CO3	3	3	1			2	1	3	3		3	2
CO4	3	3	1			3		3	2	3	3	2
CO5	3	3	1			2	2	3	2	3	3	2
		1	1- Low	Correlation	n· 2. Modera	te Correlati	on· 3_ Sub	stantial Co	rrelation	1	1	1



Effective from Session: 2020-21									
Course Code	BS 419	Title of the Course	Educational Tour	L	Т	Р	С		
Year	Ι	Semester	Ш	0	0	0	0		
Pre-Requisite	UG in Biological Science	Co-requisite							
	The main objective of this of	course is to provide the stu	idents an exposure to various research activities in t	the cou	intry an	d acqua	int		
Course Objectives	the student with state-of-the	e-art technique/instrument	s used in various research institutions and industrie	s of na	tional r	epute. T	he		
	student needs to submit a report after completion of the tour.								

Course	Course Outcomes							
CO1	Develop understanding of state-of-the-art techniques/instruments used in various reputed research institutions. and industries							
CO2	Take part in Group discussion and learn Teamwork.							
CO3	Enhance communication and social skills by communication with peers.							
CO4	Student shall be able to plan and improve the Technical Report writing skills							
CO5	Have created Interest to pursue lifelong learning.							

				Course A	Articulation N	Aatrix: (Ma	pping of C	Os with P	Os and PSOs	;)		
PO-												
PSO	PO1	PO2	PO3	PO4	PO5	PO6	PO7	PO8	PSO1	PSO2	PSO3	PSO4
CO												
CO1	3	1	1			2		3	2	3	3	3
CO2	3	2	2	1				1		3		3
CO3	3	2	2	1				1		3		3
CO4	3					2		2	2	3		3
CO5	3	2						3	2	3		3

Name & Sign of Program Coordinator	Sign & Seal of HoD